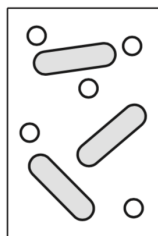
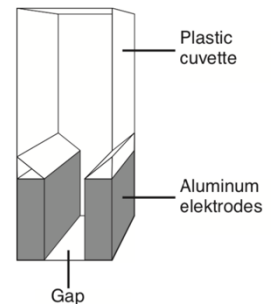
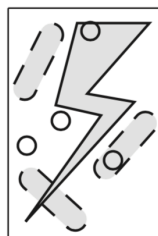


***Agrobacterium* Transformation**

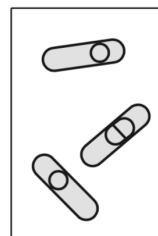
- Before started
 - Prepare cuvette
 - The cap size of the cuvette in the Soltis lab is 0.1 cm
 - Wash with deionized water
 - Put inside a baker with 70% EtOH
 - Dry inside the hood; put on top of a piece of plastic wrap
 - Wrap it and put inside 4 C (to improve transformation efficiency)
 - Pre-warm recover SOB and LB plates at 28 C
- Get the electrocompetent cell from the -80; put on ice (thaw for 10 min)
- Add 1 μ l plasmid (~100 ng) into 100 μ l agrobacterium; mix well, and put on ice
- Operate the electroporator
 - The one in the Soltis lab: Eppendorf 2510
 - Transfer the mixture to the cuvette (no bubbles! By tapping down 2X)
 - Wipe the cuvette
 - Set the voltage
 - 0.1 cm gap size: 1.8 kV
 - 0.2 cm gap size: 2.5 kV
 - Start the charging/discharging procedure
 - Pressing the PULSE key twice
 - Removing the sample



DNA and bacteria



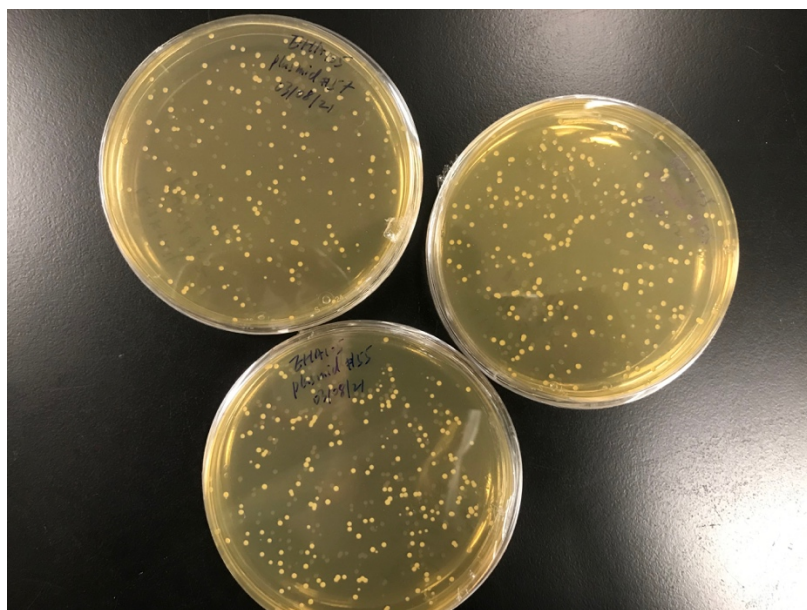
Electrical discharge
2,500 V
5 ms



DNA in bacteria

- Recovery
 - Immediately add 1 ml SOB into the cuvette and pipette up and down to mix
 - Add the mixture to the 1.5 ml centrifuge tube
- Shake in 28 C for 1-2 h
 - Put the tube inside a flask
- Use 100-200 μ l culture (~1/10 of the total volume) and spread on a LB plate with appropriate antibiotics (e.g., LB+K+Rif for strain EHA105) using glass beads
 - Wrap the plate; upside down; 28 C
 - it will take ~ 2 days

Updated on 03/12/2021



20210310_transformed.agro_plasmid55-57-75